

The Synthesis of Naturally Occurring Vitamin K and Vitamin K Analogues

Alison M. Daines, Richard J. Payne, Mark E. Humphries, and Andrew D. Abell*

Department of Chemistry, University of Canterbury, Private Bag 4800, Christchurch, New Zealand

Abstract: The synthesis of vitamin K and its analogues has been an important goal since the biochemical roles of the K vitamins were elucidated. This review presents a detailed account of syntheses of natural K vitamins and analogues that contain side chain functionality.

INTRODUCTION

A number of life's key biochemical processes, including blood coagulation, bone metabolism and cell growth, are mediated by a family of natural products referred to as the K vitamins [1]. Much work has been carried out in recent times to elucidate the mechanisms by which these compounds elicit their activity, and a number of proteins that are vitamin K-dependent have been identified in a variety of human tissues including the kidneys, spleen, lungs, uterus, placenta, thyroid, thymus, testes, and bones. As might be expected this commonality in function is reflected in a commonality in structure within this class of vitamin, i.e. they contain a naphthoquinone core, the substitution of which defines the exact K vitamin. For example, vitamin K₁ (phylloquinone) **1** has a 3-phytyl substituent while vitamin K₂ (menaquinone) **2** contains repeating unsaturated isoprene units at the 3 position. The menaquinones may be denoted as MK-n, where n is the number of repeating isoprene units (see Figure 1). The other form of vitamin K commonly encountered is 2-methyl-1,4-naphthoquinone (vitamin K₃ or menadione) **3** which lacks substitution at the 3 position. Although biologically active in vivo this compound is not found in nature.

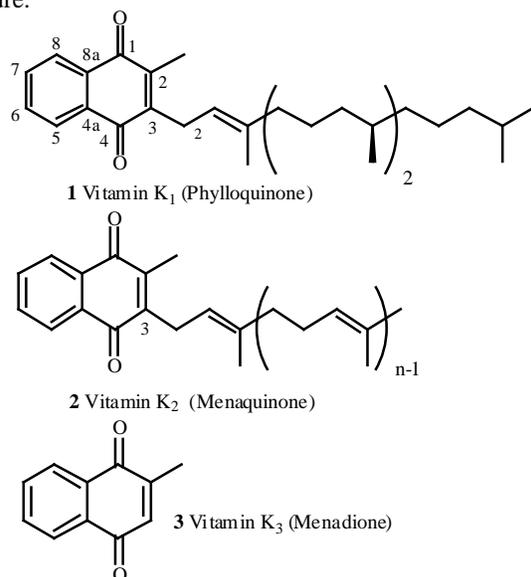


Fig. (1).

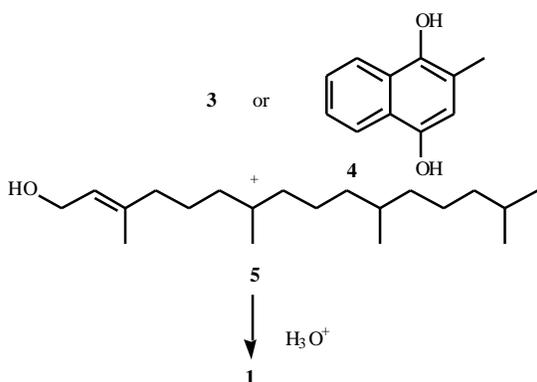
*Address correspondence to this author at the Department of Chemistry, University of Canterbury, Private Bag 4800, Christchurch, New Zealand; Tel: + 63 - 3 - 3642818; Fax: + 64 - 3 - 3642110; E-mail: a.abell@chem.canterbury.ac.nz

The process of blood coagulation has been particularly well studied and the central role of vitamin K in this process has been elucidated [1]. It is now well documented that vitamin K is essential for the proper formation of prothrombin, the inactive precursor of thrombin - the enzyme that converts fibrinogen of blood plasma into fibrin that results in blood clots. This biochemical process is itself mediated by the carboxylation of specific glutamic acid (Glu) residues to γ -carboxyglutamic acid (γ -Glu) residues in prothrombin in what is referred to as the vitamin K cycle [2]. The resulting vitamin K-dependent post-translational modification of Glu residues enables the protein to chelate calcium ions, and in the case of blood coagulation, this calcium ion is then able to form ion bridges to the phosphate head groups at the phospholipid membrane surfaces of blood platelets and endothelial cells [3]. The introduction of a γ -Glu carboxyl group is also believed to confer structural changes to the protein as a result of calcium ion mediated γ -Glu- γ -Glu cross-linking [3].

Vitamin K has also been implicated in a number of other important physiological processes including early skeletal development maintenance of bone [2,4], cellular growth [5]. In association with its role in cell proliferation, vitamin K has also been shown to inhibit the growth of several tumour cell lines [6]. It is clear that the vitamin K family is of major importance to life as we know it and as such a good deal of effort has gone into developing general and efficient methods for the preparation of the K vitamins. This work is reviewed here for the first time. In addition, these methods have been extended to allow preparation of analogues of the K vitamins to help study these and related processes.

SYNTHESIS OF NATURALLY OCCURRING K VITAMINS

As vitamin K is not synthesised by higher animals it is necessary to obtain the required daily allowance through dietary sources. In nature the K vitamins are synthesised via the shikimic acid pathway, which is also utilised in the production of the aromatic amino acids [7]. The various prenyl side chains are synthesised via the terpene biosynthetic pathway before incorporation into the final compound [8]. Vitamin K₁ is synthesised by plants and is found at significant levels in green leafy vegetables, certain legumes and some vegetable oils, such as rapeseed and soyabean oil [9]. Most fish, skeletal meats, cereals, and beverages also contain low but measurable amounts of vitamin K₁[9]. Vitamin K₂ is predominantly synthesised by anaerobic bacteria including many species that inhabit the



Scheme 1.

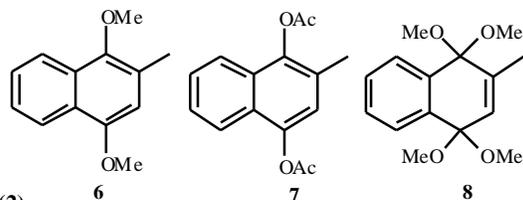
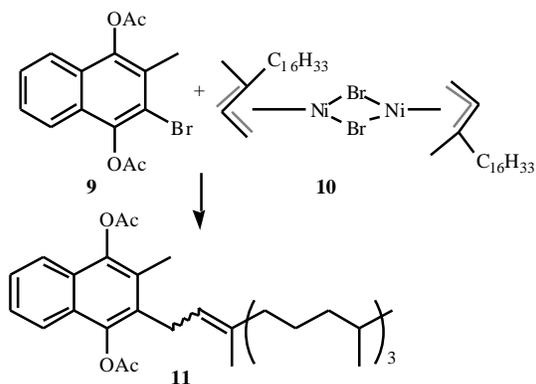


Fig. (2).

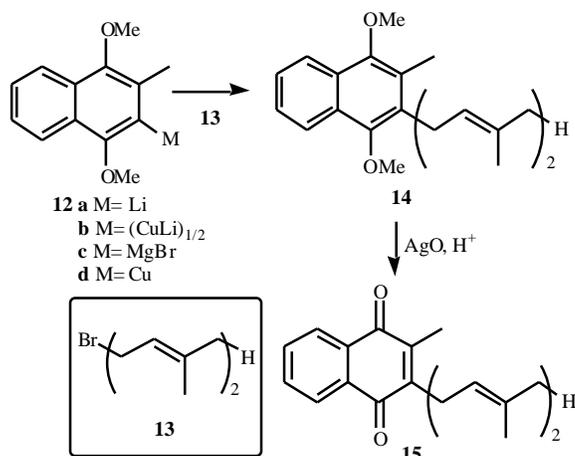


Scheme 2.

lining of the gut. The major forms of vitamin K₂ from this source are MK-10 and MK-11 produced by the *bacteroides*, MK-8 from *enterobacteria*, MK-7 from *veillonella sp* and MK-6 from *Eubacterium lentum* [9]. Vitamin K₂ is also available through the diet from animal livers and fermented foods, including cheeses [9].

Another potential source of vitamin K is via dihydrovitamin K₁, which is produced when vegetable oils are hydrogenated. Foods prepared using hydrogenated vegetable oils have shown significant quantities of dihydrovitamin K₁ [10] and after consumption of such foods, dihydrovitamin K₁ has been detected in human plasma, but its bioavailability is unknown [10]. It is estimated that in America dihydrovitamin K₁ may account for up to 30% of the daily vitamin K intake, so this may become very important in assessing vitamin K status [10].

The first reported laboratory syntheses of vitamin K₁ came independently from the laboratories of Fieser, [11] Binkley, [12] and Almquist / Klose [13] in 1939. Here, condensation of either, menadione 3 or 2-methyl-1,4-naphthohydroquinone 4, with natural phytol 5 in the presence of oxalic acid or zinc dust in acetic acid, respectively, gave vitamin K₁ (Scheme 1). Purification of the final product proved difficult and yields were low using



Scheme 3.

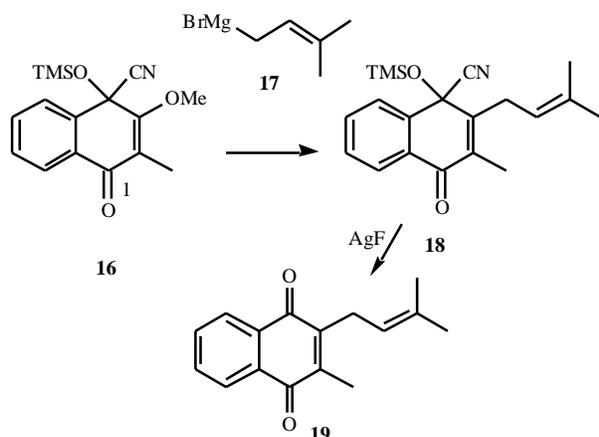
these methods, with the formation of the chromanol (undefined cyclic isomers of the corresponding hydroquinones) proving a major problem.

These early syntheses were based upon Friedel-Craft alkylations for side chain coupling, which led to mixtures of isomers at the 2 position [11-13]. For example, the conventional Friedel-Crafts coupling of natural *E*-phytol to 2-methyl-1,4-naphthohydroquinone gave, after oxidation, a mixture containing approximately 90% *E*-phyloquinone (isomerisation was occurring during the coupling process). An improvement on this method by Lindlar [14] using menadiol 1-benzoate as the starting material, and BF₃.Et₂O as the catalyst, gave complete retention of stereochemistry in the side chain. This sequence also avoided chromanol cyclisation and competing C2-alkylation, however, its use remains limited by the inherent instability of the allylic alcohol to the acidic reaction conditions [15].

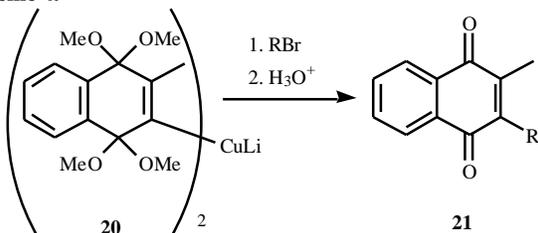
The majority of the more recent syntheses involve coupling a side chain to a suitably protected quinone core, the nature of the protection used being influenced by the nature of the subsequent chemistry and/or the deprotection methods involved. 1,4-Dimethoxynaphthalene 6, 1,4-diacetoxynaphthalene 7, and quinone bisketals 8 have all proven to be popular building blocks due to their ease of deprotection by oxidation, base or acid respectively.

The advent of transition metals in organic synthesis has greatly advanced the efficiency of coupling procedures employed for the synthesis of K vitamins. Initial reports employed allyl nickel complexes, [16] for example 2-bromo-3-methyl-1,4-diacetoxynaphthalene 9 reacts with *E*-phytylnickel complex 10 to afford the acetyl protected vitamin K₁ 11 (Scheme 2). The *Z*/*E* ratio of the products obtained was found to vary greatly with reaction solvent; *N*-methylpyrrolidone giving the most desirable results, a 8:2 ratio in favour of the *E* isomer (75% combined yield). Vitamin K₂ was synthesised in an analogous manner in 85% yield and in a ratio of 7:3 favouring of the *E* isomer. The *Z*/*E* ratio was increased to 87:13, in this example, by low temperature recrystallisation.

Snyder and Rappaport reported a synthesis of a series of menaquinones from a 2-metallo-3-methyl-1,4-dimethoxynaphthalene (12) and a suitable aldehyde or alkyl halide (Scheme 3) [17]. Of the metallonaphthalenes investigated,



Scheme 4.

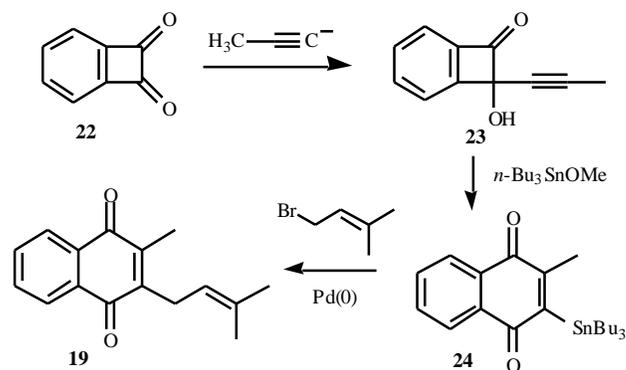


Scheme 5.

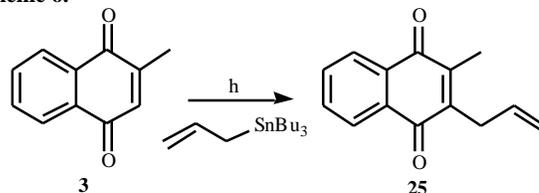
Grignard reagents proved the most reliable, giving excellent retention of the *E* configuration (98-99%). Coupling the Grignard reagent of 2-bromo-3-methyl-1,4-dimethoxynaphthalene (**12c**) with geranyl bromide (**13**) gave MK-2 in 80%, while solanesyl bromide afforded MK-9 in 73% yield. In both cases 2-bromo-3-methyl-1,4-dimethoxynaphthalene was observed as a contaminating by product.

Evans and Hoffman published a regioselective isoprenylation of benzo- and naphthoquinones as a route to the vitamin K family [18]. Cyanide-catalysed addition of trimethylsilyl cyanide (TMSCN) to 2-methoxy-3-methyl-1,4-naphthoquinone gave the protected quinone **16**. Quinone **16** then underwent C1 carbonyl addition with prenyl magnesium bromide **17** followed by a Cope rearrangement to the protected MK-1 **18**. Deprotection with AgF afforded the desired quinone **19** in 71% yield (Scheme 4).

Organocuprates have been used for the synthesis of the K vitamins. Chenard *et al.* [19] prepared menaquinone and phylloquinone from cuprates of quinone bisketals with allyl halides. The cuprate was itself prepared by metalation of the corresponding bromide. High yields and good stereoselectivity at the ² position of the K vitamin product **21** were obtained (Scheme 5). The cuprate activity is substrate dependent with two alkyl groups being transferred with groups such as allyl bromide and one group being transferred with bulkier substrates such as benzyl chloride, cyclohexanecarbonyl acid chloride, and benzyl bromide [19]. Syper *et al.* [20] also utilised cuprate chemistry in the coupling of prenyl bromide and geranyl bromide to 2-bromo-3-methyl-1,4-dimethoxynaphthalene to give the protected forms of MK-1 and MK-2 in 68% and 78% respectively. A method for the preparation of MK-4 was also reported by Garcias *et al.* [21] in 1994. This method involves coupling 2-bromo-3-methyl-1,4-dimethoxynaphthalene with allylic



Scheme 6.



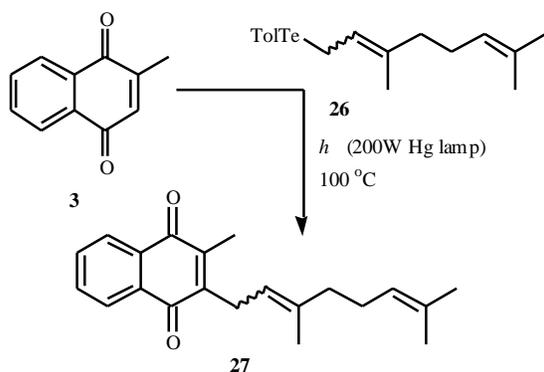
Scheme 7.

aldehydes using *n*BuLi to give the corresponding benzylic alcohol. Removal of the hydroxyl group under Birch conditions was shown to proceed with complete retention of ² stereochemistry. A final oxidative deprotection of the quinone, gave MK-1 and MK-3 in 35% and 30% yield respectively.

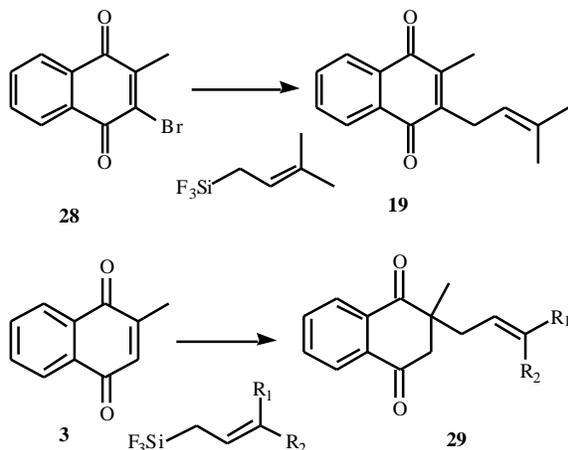
Up until the early 1990's the majority of syntheses of K vitamins involved either Grignard reagents, organolithiums or organocuprates as discussed above. These methods required the use of protected quinones to suppress competing quinone reduction. The introduction of softer nucleophiles allowed direct allylation of the quinone nucleus without the need for protection. This allowed allylsilanes and allylstannanes to be used for direct allylation of unprotected quinones. For example, MK-1 **19** has been synthesised from stannylquinones which are themselves derived from alkenylcyclobutenones [22]. Addition of a propyne nucleophile to dione **22** followed by stannylation with *n*-Bu₃SnOMe and subsequent Liebeskind-Moore rearrangement afforded the stannylquinone **24** in 49% yield over the 2 steps. A final Stille coupling with prenyl bromide gave MK-1 in 90% yield (Scheme 6).

Allylstannanes have also been photochemically coupled to benzo- and naphthoquinones as outlined in Scheme 7 [23a]. The reaction, performed under 315nm light, in benzene or acetonitrile, gave a mixture of products from which 2-allyl-3-methyl-1,4-naphthoquinone **25** crystallised in 15-40% yield. This approach avoids the use of more traditional Lewis acid catalysis for substrate activation. A related route to vitamin K based on a photochemical-mediated radical coupling of a geranyl organotellurium species to **3** was reported in 2002 (Scheme 8) [23b]. Thus, geranyl tolyl telluride **26** (73:27 mixture of *E* and *Z* isomers, prepared from the corresponding bromide on treatment with SmI₂ and ditolyl ditelluride) was coupled to **3** to give MK-2 (**27**) in 73% yield (7:3 *E* to *Z*).

In 1995 a synthesis of MK-1 was reported by Hagiwara *et al.* [24] based upon a regioselective allylation of 2-bromo-3-methyl-1,4-naphthoquinone **28** with (3-methyl-2-



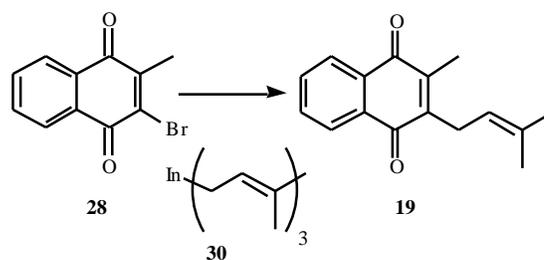
Scheme 8.



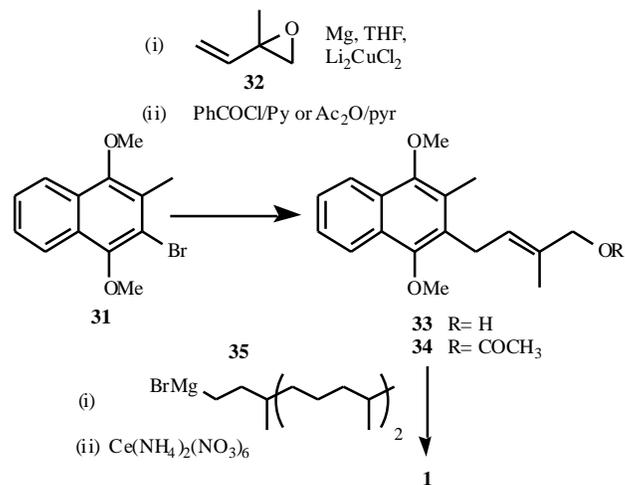
Scheme 9.

butenyl)trifluorosilane in formamide at 80 °C (Scheme 9). By contrast, the alkylation of the non-halogenated 2-methyl-1,4-naphthoquinone **3** occurred at the substituted C2 carbon to give **29**. An allylindium **30** has also been used to prepare simple menaquinones from 2-bromo-3-methyl-1,4-naphthoquinone with the indium transferring two of the three available allyl groups from the complex [25]. Prenylindium **30** and **28** react to give the MK-1 vitamin in 67% yield (see Scheme 10). The authors postulate that the reaction proceeds by addition to the carbonyl group followed by a [3,3] sigmatropic rearrangement of the prenyl group, rather than the direct substitution at the 2-position of the ring. MK-1 has also been prepared via the coupling of 2-methyl-1,4-naphthoquinone to prenyl units using a lead bromide and aluminium powder catalyst. This catalyst allows the coupling to be carried out at room temperature under conditions that are not strictly anhydrous. The mono-prenylated product MK-1 was obtained exclusively in 50% yield [26].

An alternative approach to the K vitamins involves the stepwise elongation of a suitable short side chain analogue. This methodology has been used to prepare natural phyloquinone (Vitamin K₁) as a mixture of stereoisomers [27]. The addition of the Grignard reagent derived from 2-bromo-3-methyl-1,4-dimethoxynaphthalene **31**, to isoprene epoxide **32**, gave the short chain alcohol derivative **33**. Conversion of **33** into the corresponding acetate **34**, followed by addition of a second Grignard reagent gave vitamin K₁ (Scheme 11).



Scheme 10.

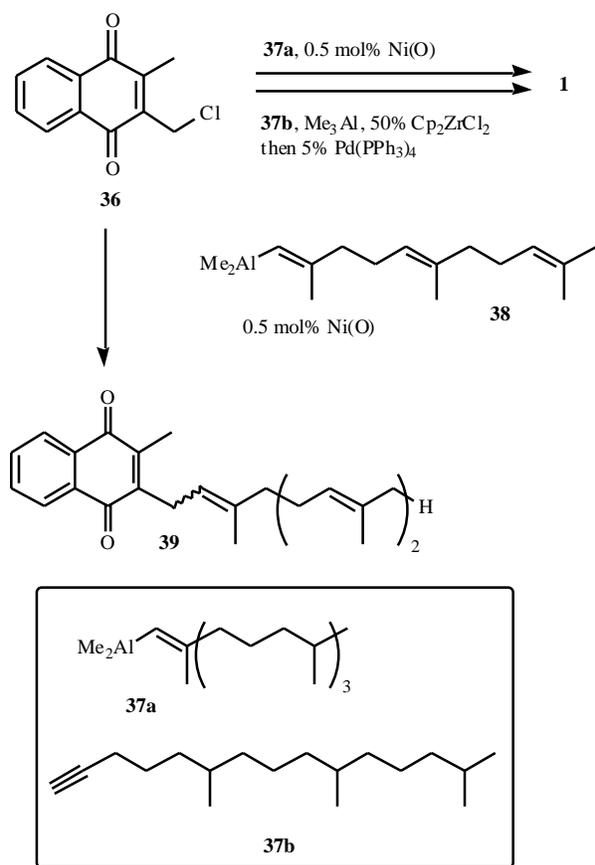


Scheme 11.

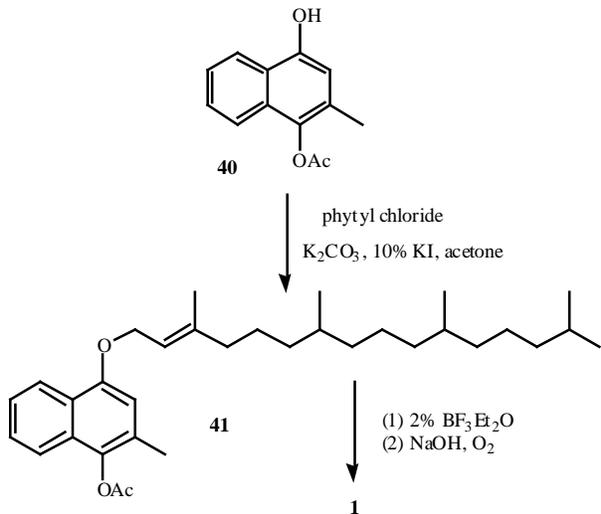
More recently, Ni(0) catalysts have been used by Lipshutz to extend the side chains of simple derivatives to provide a convenient synthesis of phyloquinone and menaquinones (Scheme 12) [28a]. Here, chloromethylated quinone (**36** (prepared either, by direct chloromethylation of the quinone or from oxidation of corresponding benzylic chloride)) was reacted with the trimethylaluminium functionalised side-chains **37a** and **38** in the presence of 0.5 mol% Ni(0) to give racemic phyloquinone **1** in 88% yield and MK-3 **39** in 93% yield, respectively [28a]. The chloromethyl quinone **36** has also been used to prepare vitamin K via direct Pd-catalysed alkenylation with an alkenylalane derived from **37b**, Scheme 12 [28b].

There are a few other isolated reports on the synthesis of the vitamin K family. For example, α -cyclodextrin has been shown to catalyse a key coupling step in the synthesis of vitamin K₁, K₂, and analogues [29]. The reaction of 2-methyl-1,4-naphthoquinone and a respective bromide using α -cyclodextrin in phosphate buffer containing 30% methanol gave the K vitamin in yields ranging from 40-60%. It was postulated that cyclodextrin acts as a ligase and/or oxygenase, i.e. a 'vitamin K synthase', in this reaction.

In another isolated report, natural phyloquinone **1** has been synthesised by an *O*-alkylation of the menadiol monoacetate **40** giving ether **41** which is followed by a Lewis acid catalysed rearrangement [27]. The intramolecular rearrangement of **41** occurs under mild reaction conditions and low catalyst loading to give a high (*E*)-selectivity in the double bond formation (Scheme 13). This approach has received little attention.

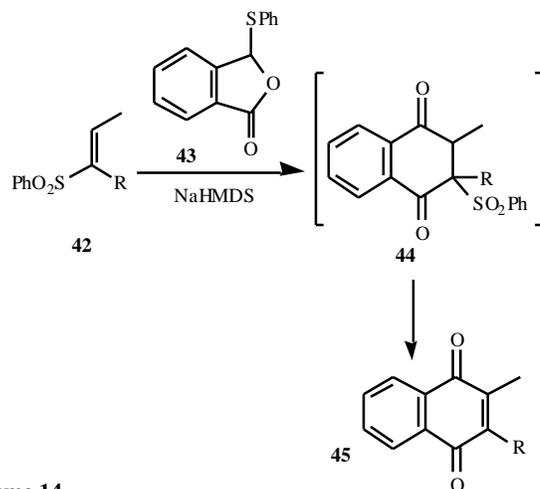


Scheme 12.

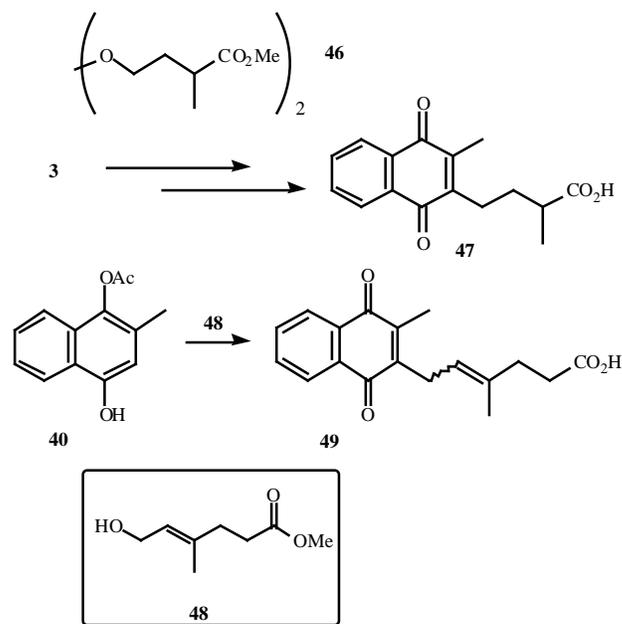


Scheme 13.

In 1995 Tso *et al.* [30] published a novel one-pot procedure to K vitamins that is conceptually quite different to those already discussed (Scheme 14). Treatment of 3-substituted isobenzofuranone **43** with base generated a quinone methide which underwent a [4+2] cycloaddition with the alkyl phenyl-sulfone **42**. Elimination of the sulfone from intermediate **44** *in situ* then gave phyloquinone in 61% yield. MK-1, MK-2 and MK-9 were also prepared in this manner in 60%, 63%, and 64% yield, respectively [30].



Scheme 14.

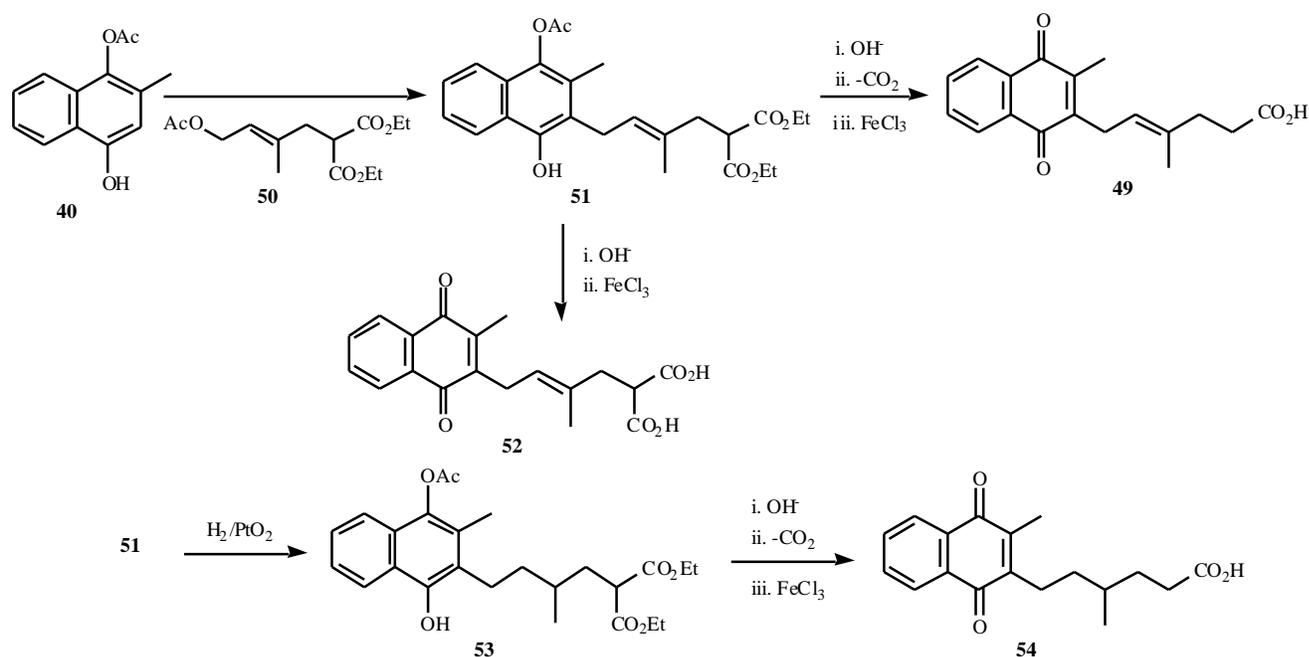


Scheme 15.

Synthesis of vitamin K analogues with side chain functionality

The development of methods for the synthesis of analogues of vitamin K have also received a good deal of attention with a view to understanding structure activity relationships (SAR) within this family of compounds. In addition, vitamin K analogues are of interest as potential inhibitors of vitamin K dependant carboxylase and vitamin K epoxide reductase [31,32]. They have also been used to investigate and characterise possible metabolites of the K vitamins.

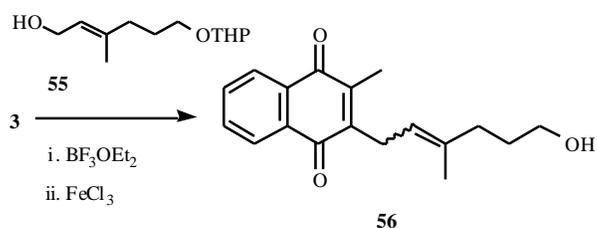
The stereochemistry at the side chain alkene of vitamin K₁ is known to be essential for biological activity [33]. The natural form of vitamin K₁ and the MK's has a *E* configuration, and the synthetic *Z* isomer exhibits little or no activity [33,34]. Of the menaquinone series MK-3 to MK-5 appear to be the most biologically active *in vivo*. [7]. The longer chain menaquinones have been shown to be highly active *in vitro* but they may be less well absorbed, due to

**Scheme 16.**

increased hydrophobicity, leading to a lower biological activity [35]. The unnatural vitamin K_3 has also been shown to exhibit biological activity but it is unknown if this is alkylated *in vivo* to produce an active compound [7]. An investigation of the biological activity of a series of analogues of vitamin K has also provided some structure activity information on these compounds [7]. 2-Methyl-4-amino-1-naphthol and 2-methyl-1-naphthol were shown to have biological activity similar to menadione [7]. Diphosphate, disulfate, diacetate, and dibenzoate forms of the reduced K series exhibited full biological activity, [7] possibly due to the fact that it is the reduced form of vitamin K that is biologically active. A methyl group at the 2 position is essential for activity, with the 2-ethyl and other alkylated derivatives showing no activity [33,34]. In addition to this the 2-chloro and 2-bromo derivatives are both potent antagonists of vitamin K [7]. 2-Phenyl-1,4-naphthoquinone has been shown to exhibit biological activity, however it is thought to be methylated *in vivo* to vitamin K_1 [7]. Saturation of the double bond of vitamin K_1 reduces activity considerably, but the compound still exhibits more activity than branched chain K_1 analogues [7]. Finally, alteration of the naphthoquinone core in any manner also leads to a loss in biological activity [7].

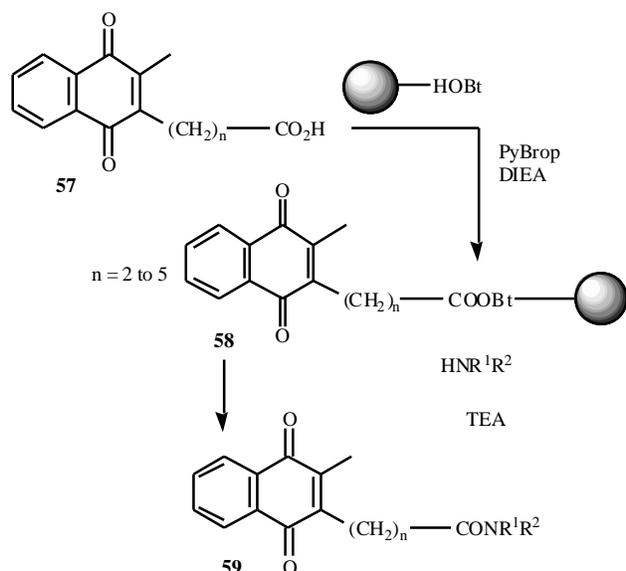
A wide variety of analogues of vitamin K have been synthesised including substituted vitamin K epoxides, [31,32] methylated naphthoquinones, [36] and side-chain functionalised vitamin K analogues [37-48] since the early efforts in the mid 1970's. In this review we are concerned solely with the last class of analogue since the chemistry involved in their synthesis correlates with that already discussed.

One of the earliest syntheses of vitamin K analogues (see 47 and 49, Scheme 15) was reported by Watanabe *et al.* [39]. Reaction of the radical derived by thermolysis of peroxide 46

**Scheme 17.**

with 2-methyl-1,4-naphthoquinone 3, followed by ester hydrolysis, gave the vitamin K analogue 47 in 50% yield based on the peroxide. However, it is important to note that the starting peroxide was prepared in a three-step synthesis in only 5% yield. The authors also reported condensation of 1-acetoxy-4-hydroxy-2-methylnaphthalene 40 with methyl 6-hydroxy-4-methylhex-4-enoate 48 to give 49 in 42% yield to give a 3:1 mixture of *Z:E* isomers.

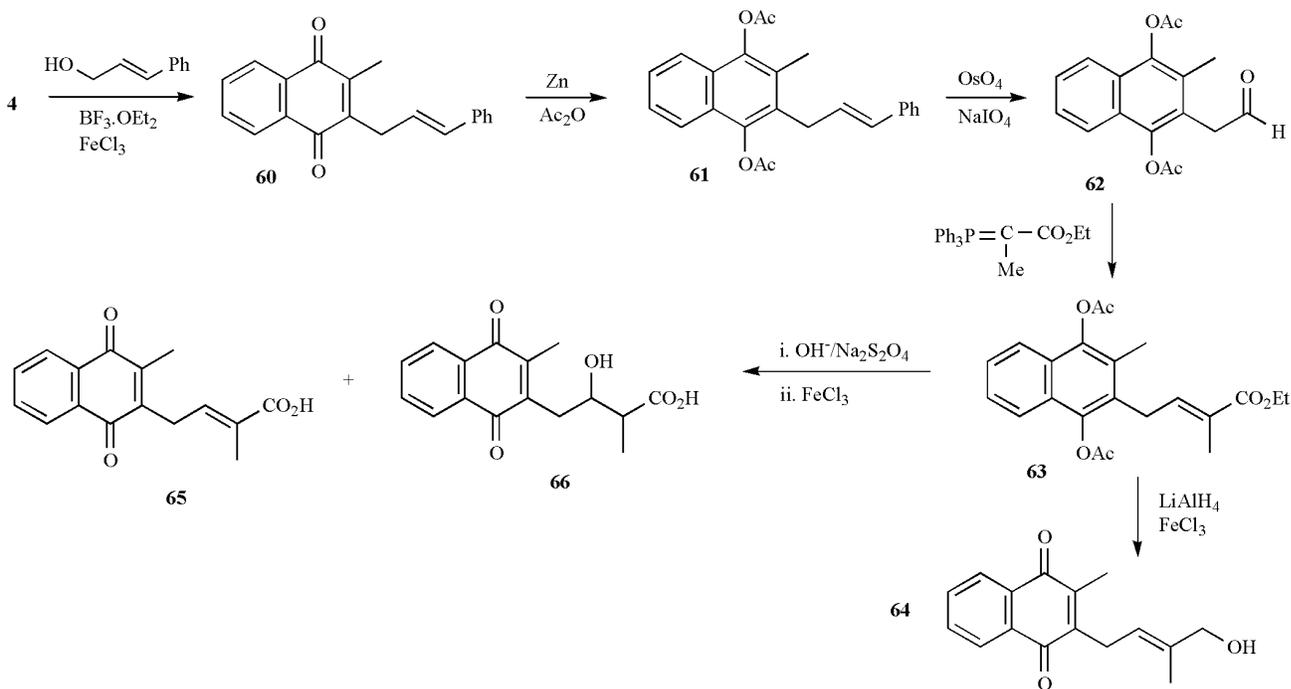
Watanabe also reported [40] a related synthetic approach to carboxylated side-chain vitamin K analogues (Scheme 16). Condensation of monoacetate 40 and diester 50 in the presence of $\text{BF}_3 \cdot \text{Et}_2\text{O}$ afforded the malonate intermediate 51 in 66% yield. As before, this synthesis is hindered by the multi-step preparation of compound 50. Intermediate 51 however, proved to be a useful synthetic intermediate being able to be hydrolysed to the unsaturated diacid 52, decarboxylated to the unsaturated monoacid 49 or hydrogenated and hydrolysed to the saturated acid 54. In a related sequence (Scheme 17) the alcohol derivative 56 was prepared from the 2-methyl-1,4-naphthoquinone 3 by coupling the monoprotected diol 55 to the quinone in the presence of $\text{BF}_3 \cdot \text{Et}_2\text{O}$. Deprotection and oxidation afforded the analogue 56 in 29% yield as a mixture of *Z/E* isomers. More recently, saturated naphthoquinone acids 57 (c.f. 54 in Scheme 17) have been used to prepare a library of 1,4-naphthoquinone



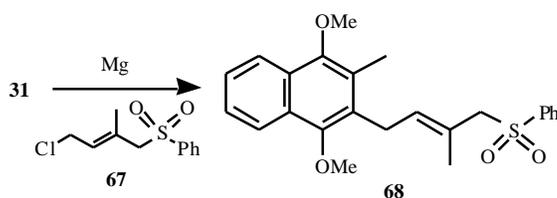
Scheme 18.

amides **59** which were screened for inhibitory activity against Trypanothione Reductase from *Trypanosoma cruzi*, the causal agent of Chagas disease, Scheme 18 [49].

A series of vitamin K analogues bearing an alcohol functionality have been prepared as outlined in Scheme 19 [38]. Condensation of 2-methyl-1,4-naphthohydroquinone **4** with cinnamoyl alcohol followed by oxidation with ferric chloride afforded the phenylpropenylquinone **60**. Reductive acetylation then gave the acetyl protected hydroquinone **61** which was oxidatively cleaved with $\text{OsO}_4 / \text{NaIO}_4$ to give the corresponding aldehyde **62**. Wittig chemistry was then used to extend the side chain to give **63** which was either, reduced with lithium aluminium hydride to give alcohol **64**,



Scheme 19.

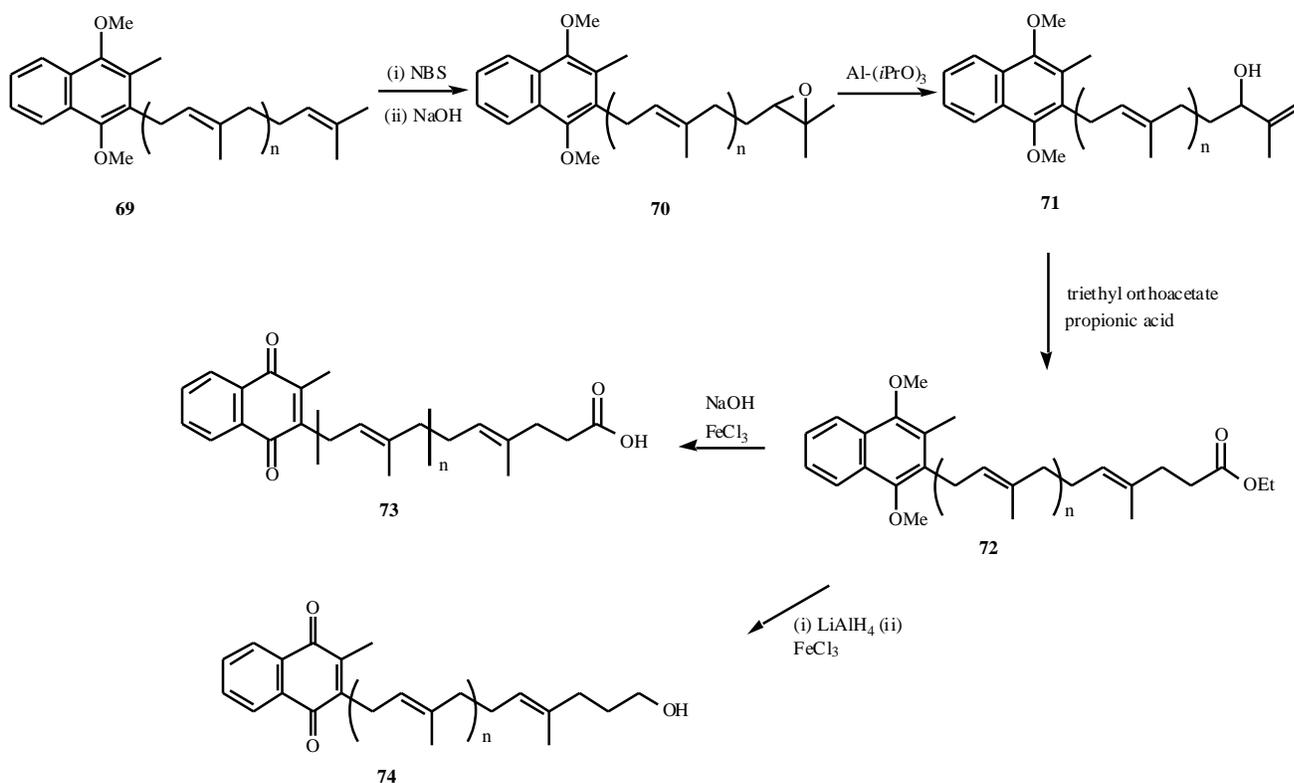


Scheme 20.

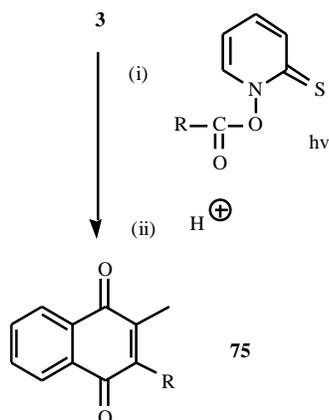
or hydrolysed and oxidised to give a mixture of acids **65** and **66** [38].

An efficient synthesis of sulfone functionalised analogues of vitamin K has been reported by Fujita *et al.* (Scheme 20) [41]. This was achieved via the Grignard coupling of 2-bromo-3-methyl-1,4-dimethoxynaphthalene **31** with *E*-4-chloro-2-methyl-1-phenylsulfonyl-2-butene **67** to give sulfone analogue **68** in 82% yield. Sulfone **67** was conveniently obtained from the reaction of isoprene with benzenesulfonyl chloride in the presence of copper chloride/TEA hydrochloride as catalyst.

An important study by Terao *et al.* [42] reports the synthesis of a series of quinone derivatives that contain modified polyprenyl side chains (Scheme 21). The regio-specific modification of the terminal prenyl group of ubiquinone, menaquinone or tocoquinone side chains gave a convenient synthesis of quinone acids, quinone amides, quinone alcohols and quinone methylketones. In this work, isomerisation of the epoxide **70** gave the allylic alcohol **71** which underwent a Claisen rearrangement (irrespective of the number *n* of prenyl units in the side chain) to the ester **72**. This was followed by either, deprotection and oxidation to give the acid **73**, or reduction to give the alcohol **74**. The result of the sequence is to increase the length of the side-chain by two carbon units as well as introducing terminal functionality.



Scheme 21.



Scheme 22.

Work has been reported by Barton *et al.* on the synthesis of hindered quinones via radical reactions, Scheme 22 [43]. On photolysis with tungsten light, acyl derivatives of *N*-hydroxy-2-thiopyridone afford carbon radicals that readily add to 2-methylnaphthoquinone **3**. The resulting adducts then provide access to a number of hindered naphthoquinones including vitamin K based compounds **75** with side chain functionality such as phenyl, isopropyl, and carboxyethyl.

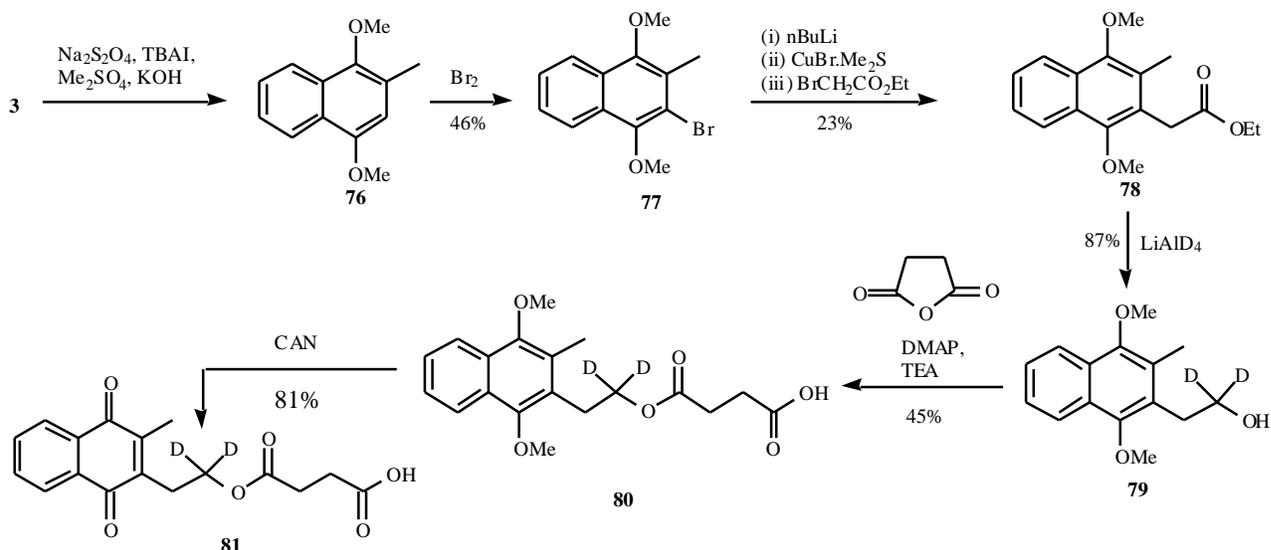
The synthesis of a deuterated analogue of vitamin K was published in 2002 by Payne and Abell, where a terminal carboxylic acid was incorporated into the structure to allow attachment to a protein via lysine side chain coupling (Scheme 23) [44]. The synthesis began with a reductive methylation of **3** using dimethyl sulfate and sodium

dithionite in the presence of the phase-transfer catalyst, tetrabutylammonium iodide (TBAI), to give the methoxy protected naphthoquinone **76**. This was then brominated to afford the aryl bromide **77**. The introduction of the C3 substituent was then achieved by a cuprate-mediated coupling of ethyl bromoacetate to a lithiated derivative of **77**, to give **78**. The ester **78** was then reduced with lithium aluminium deuteride to give the alcohol **79**, labelled with deuterium at the 2-position of the hydroxyethyl group. Introduction of the side chain linker was achieved by reaction with succinic anhydride in the presence of dimethylaminopyridine (DMAP) to give the protected vitamin K analogue **80** in 45% yield. Finally, oxidative deprotection of **80** with ceric ammonium nitrate (CAN) afforded the desired vitamin K analogue **81** in 81% yield, which was then used in conjugation studies to lysozyme [44].

Finally, a number of thioalkyl vitamin K analogues have also been reported which show interesting medicinal properties, including antitumour activity [45,46]. The general synthetic scheme for production of these analogues involves the direct reaction of menadione with the respective thiol in a mixture of methanol and 2-propanol. A number of compounds of this type are currently under investigation as potential anticancer compounds [47,48].

CONCLUSION

The vitamin K family of compounds play a central role in a number of biochemical processes that are fundamental to life. As such their synthesis has attracted a good deal of attention and the methods developed now provide access to non-natural analogues that are of medicinal interest. Early



Scheme 23.

Friedel-Craft based syntheses suffered from problems of low yield and isomerisation of the side chain due to harsh reaction conditions employed. Much milder methods are now available that simplify the need for protection of the naphthoquinone core while being compatible with a range of functionality.

REFERENCES:

- Nelsetuen, G. L.; Shah, A. M.; Harvey, S. B. *Vitamins and Hormones* **2000**, *58*, 355.
- Dowd, P.; Ham, S. W.; Geib, S. J. *J. Am. Chem. Soc.* **1991**, *113*, 7734.
- Dowd, P.; Hershline, R.; Ham, S. W.; Naganathan, S. *Science* **1995**, *269*, 1684. Berkner, K. L.; Pudota, B. N. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 466. Houben, R. J. T. J.; Jin, D.; Stafford, D. W.; Proost, P.; Ebberink, R. H. M.; Vermeer, C.; Soute, B. A. M. *Biochem. J.* **1999**, *341*, 265.
- Vermeer, C.; Jie, K. S. G.; Knapen, M. H. J. *Ann. Rev. Nutr.* **1995**, *15*, 1. Shearer, M. J.; Bach, A.; Kohlmeier, M. *J. Nutr.* **1996**, *126*, 1181s. Shearer, M. J. *Proc. Nutr. Soc.* **1997**, *56*, 915.
- Nakano, T.; Kawamoto, K.; Kishino, J.; Nomura, K.; Higashino, K.; Arita, H. *Biochem. J.* **1997**, *323*, 387.
- Prasad, K. N.; Edwards-Prasad, J.; Sakamoto, A. *Life Sci.* **1981**, *29*, 1387. Ngo, E. O.; Sun, T.-P.; Chang, J.-Y.; Wang, C.-C.; Chi, K.-H.; Cheng, A.-L.; Nutter, L. M. *Biochem. Pharm.* **1991**, *42*, 1961. Ni, R.; Nishikawa, Y.; Carr, B. I. *J. Biol. Chem.* **1998**, *273*, 9906.
- Suttie, J. W. In *Handbook of vitamins*; Machlin, L. J., Ed.; Marcel Dekker Inc: New York, **1991**, p 145-194.
- Begley, T. P.; Kinsland, C.; Taylor, S.; Tandon, M.; Nicewonger, R.; Wu, M.; Chiu, H.-J.; Kelleher, N.; Campobasso, N.; Zhang, Y. *Topics Curr. Chem.* **1998**, *195*, 93.
- Shearer, M. J. *Lancet* **1995**, *345*, 229.
- Booth, S. L.; Pennington, J. A. T.; Sadowski, J. A. *Lipids* **1996**, *31*, 715.
- Fieser, L. F. *J. Am. Chem. Soc.* **1939**, *61*, 2559.
- Binkley, S. B.; Cheney, L. C.; Holcomb, W. F.; McKee, R. W.; Thayer, S. A.; MacCorquodale, D. W.; Doisy, E. A. *J. Am. Chem. Soc.* **1939**, *61*, 2558.
- Almquist, H. J.; Klose, A. A. *J. Am. Chem. Soc.* **1939**, *61*, 2557.
- Lindlar, H. Patent No. 320582, Switzerland, 1953.
- Stevens, K. L.; Jurd, L.; Manners, G. *Tetrahedron* **1972**, *28*, 1939.
- Sato, K.; Inoue, S.; Saito, K. *J. Chem. Soc. Perkin Trans. 1* **1973**, 2289.
- Snyder, C. D.; Rappaport, H. *J. Am. Chem. Soc.* **1974**, *96*, 8046.
- Evans, D. A.; Hoffman, J. M. *J. Am. Chem. Soc.* **1976**, *98*, 1983.
- Chenard, B. L.; Manning, M. J.; Reynolds, P. W.; Swenton, J. S. *J. Org. Chem.* **1980**, *45*, 378.
- Syper, L.; Kloc, K.; Mlochowski, J. *Tetrahedron* **1980**, *36*, 123.
- Garcias, X.; Ballester, P.; Capo, M.; Saa, J. M. *J. Org. Chem.* **1994**, *59*, 5093.
- Liebeskind, L. S.; Foster, B. S. *J. Am. Chem. Soc.* **1990**, *112*, 8612.
- (a) Maruyama, K.; Imahori, H. *Bull. Chem. Soc. Jpn.* **1989**, *62*, 816.
(b) Yamago, S.; Hashidume, M.; Jun-ichi, Y. *Tetrahedron* **2002**, *58*, 6805.
- Hagiwara, E.; Hatanaka, Y.; Gohda, K.-I.; Hiyama, T. *Tetrahedron Lett.* **1995**, *36*, 2773.
- Araki, S.; Katsumura, N.; Butsgan, Y. *J. Organomet. Chem.* **1991**, *415*, 7.
- Khanna, R. N.; Singh, P. K. *Synth. Commun.* **1990**, *20*, 1743.
- Schmid, R.; Antoulas, S.; Ruttimann, A.; Schmid, M.; Vecchi, M.; Weiser, H. *Helv. Chem. Acta* **1990**, *73*, 1276.
- (a) Lipshutz, B. H.; Kim, S.-K.; Mollard, P.; Stevens, K. L. *Tetrahedron* **1998**, *54*, 1241.
(b) Huo, S.; Negishi, E.-I. *Org. Lett.* **2001**, *3*, 3253.
- Tabushi, I.; Yamamura, K.; Fujita, K.; Kawakubo, H. *J. Am. Chem. Soc.* **1979**, *101*, 1019.
- Tso, H.-H.; Chen, Y.-J. *J. Chem. Res. (S)* **1995**, 104.
- Silverman, R. B.; Oliver, J. S. *J. Med. Chem.* **1989**, *32*, 2138.
- Ryall, R. P.; Nandi, D. L.; Silverman, R. B. *J. Med. Chem.* **1990**, *33*, 1791.
- Shearer, M. J. In *Encyclopaedia of Food Science, Food Technology and Nutrition*; Macrae, R.; Robinson, R. K.; Sadler, M. J., Eds.; Academic Press: London, **1993**, p 4804.
- Suttie, J. W. *Ann. Rev. Biochem.* **1985**, *54*, 459.
- Suttie, J. W. *Ann. Rev. Nutr.* **1995**, *15*, 399.
- Schmid, R.; Goebel, F.; Warnecke, A.; Labahn, A. *J. Chem. Soc. Perkins Trans. 2* **1999**, 1199.
- Masaki, Y.; Sakuma, K.; Kaji, K. *Chem. Pharm. Bull.* **1985**, *33*, 1930.
- Okamoto, K.; Watanabe, M.; Kawada, M.; Goto, G.; Ashida, Y.; Oda, K.; Yajima, A.; Imada, I.; Morimoto, H. *Chem. Pharm. Bull.* **1982**, *30*, 2797.
- Watanabe, M.; Kawada, M.; Nishikawa, M.; Imada, I.; Morimoto, H. *Chem. Pharm. Bull.* **1974**, *22*, 566.
- Watanabe, M.; Okamoto, K.; Imada, I.; Morimoto, H. *Chem. Pharm. Bull.* **1978**, *26*, 774.

- [41] Fujita, Y.; Ishiguro, M.; Onishi, T.; Nishida, T. *Synthesis* **1981**, 469.
- [42] Terao, S.; Shiraisi, M.; Kato, K.; Ohakawa, S.; Ashida, Y.; Maki, Y. *J. Chem. Soc. Perkin Trans. 1* **1982**, 2909.
- [43] Barton, D. H. R.; Sas, W. *Tetrahedron* **1990**, *46*, 3419.
- [44] Payne, R. J.; Abell, A. D. *Chem. N. Z.* **2002**, *66(1)*, 16.
- [45] Nishikawa, Y.; Carr, B. I.; Wang, M.; Kar, S.; Finn, F.; Dowd, P.; Zheng, Z. B.; Kerns, J.; Naganathan, S. *J. Biol. Chem.* **1995**, *270*, 28304. Carr, B. I.; Wilcox, C. S.; Kerns, J. K. *PCT Int. Appl.* **2000**, 08495.
- [46] Kar, S.; Adachi, T.; Carr, B. I. *J. Cell Phys.* **2002**, *190*, 356.
- [47] Ni, R.; Nishikawa, Y.; Carr, B. I. *J. Biol. Chem.*, **1998**, *273*, 9906. Tamura, K.; Southwick, E. C.; Kerns, J.; Rosi, K.; Carr, B. I.; Wilcox, C.; Lazo, J. S. *Cancer Research*, **2000**, *60*, 1317. Wang, Z.; Nishikawa, Y.; Wang, M.; Carr, B. I. *Journal of Hepatology* **2002**, *36*, 85.
- [48] Chen, C.; Liu, Y.; Shia, K.; Tseng, H. *Biorg. Med. Chem. Lett.* **2002**, *12*, 2729.
- [49] Salmon-Chemin, L.; Lemaire, A.; De Freitas, S.; Deprez, B.; Sergheraert, C.; Davioud-Charvet, E. *Bioog. Med. Chem. Lett.* **2000**, *10*, 631.